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GREEN SYNTHESIS OF NANOPARTICLES AND THEIR EFFECT ON PLANT GROWTH AND DEVELOPMENT: A REVIEW

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ABSTRACT

Nanotechnology permits advance research in several areas and opens a large scope in the field of biotechnology and agriculture industry due to unique physiochemical properties. To fulfill the increasing demand of world population higher agricultural productivity is needed to boost the yield. This review presents the current literature and key role of nanoparticles on plant growth, development and yield. The synthesis of nanoparticles by green method with the use of plant extract which is nontoxic, cost effective, ecofriendly over physical and chemical methods. Phytochemical constituents in the plant extract such as phenols, proteins, flavonoids, carbohydrates, alkaloids and amino acids is responsible for the reduction of size of nanoparticles.

Keywords : Nanoparticles, Green method, Phenols, Proteins, Flavonoids, Carbohydrates, Alkaloids and Amino acids

Introduction

In science and technology, huge amount of development has been brought by nanotechnology in the recent years (Kalpana and Rajeswari, 2018). Nanotechnology brings revolution in all over the world (Arora et al., 2012).Particles within the size range of 1-100 nm are considered as nanoparticles (NPs) (Chung et al., 2016). Important properties of NPs have a large surface area, high surface energy and quantum confinement (Nel et al., 2006). It has become important to increase crop production to feed the growing world population, to meet the increasing demand. Nanotechnology, in the current technological innovations, obtained an important position in transforming agriculture and food production (Nair et al., 2010). Although fertilizers are very important for plant growth and development, most of the applied fertilizers are rendered unavailable to plants due to a variety of factors such as leaching, photolysis, hydrolysis, and decomposition (Siddiqui et al., 2015). Thus, it is critical to advance research in order to minimize nutrient losses in fertilization and increase crop yield through the use of new applications enabled by nanomaterials and nanotechnology (Singh et al., 2015). Nanotechnology has enormous potential in terms of high reactivity, agricultural uprising, improved bioavailability, bioactivity, and NPs surface effects (Gutierrez et al., 2011). Engineered nanoparticles have the ability to enter plant cells and leaves, as well as transport DNA and chemicals into plant cells (Galbraith 2007 and Torney et al.,

2007).Nanomaterials have ability to engineer plant function but the mode of transport, absorption and distribution of nanoparticles within plant word is still remaining poorly understood. Nanobionics to engineer plant function opens a gate in the new research field at the interface of nanotechnology and plant biology (Giraldo *et al.*, 2014).

Harmful impact of solvents and synthetic reactants on environment due to intensive use, for this reason need an alternative 'green' method application which is environment friendly reactants for the preparation of nanomaterials (Leon *et al.*, 2013). In the synthesis of NPs medicinal plants were preferably choose that already reported for biomedical properties and having wide range of natural products (Kumar and Kumar, 2017). Bioactive phytochemicals constituents reacted to reduce metals into metal oxide and showing good stability in the formation of NPs (Mishra and Sharma, 2015).

Green synthesis of Nanoparticles

The method generally used for producing nanoparticles is top down approach and bottom up approach (Sepeur, 2008). In top down approach various physical and chemical methods are used for size reduction of material to producing nanoparticles (Meyers *et al.*, 2006). The major limitation of this method is the imperfection of the surface structure of nanoparticles because physical properties and surface chemistry are highly dependent on surface structure (Thakkar *et al.* 2010). In Bottom up approach, small entities are joined e.g. molecules, atoms and small particles of nanometer range by using chemical and biological method (Mukherjee *et al.*, 2001). Biological method, synthesis of nanoparticles involved microorganisms, plants or plant extract and enzyme, it is more ecofrienldy over physical and chemical methods (Nair *et al.* 2002 and Schultz *et al.*, 2000). Plant or plant extract reduces the complexity of maintaining cell culture of

biological method for biosynthesis of nanoparticles (Wilner *et al.*, 2006). Biological entities carry capping and stabilizing agents. Enzymes, sugar, proteins and phytochemicals like flavonoids, terpenoids, phenolics, cofactors acts as stabilizing and reducing agents (Kaushik *et al.*, 2010 and Khiarissova *et al.*, 2013 Figure 1& Table1).



Fig. 1: Schematic representation of various methods for synthesizing nanoparticles

Table 1 : S	ynthesis	of nanopa	rticles via	different	plant parts
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Plant	Biosynthesis of nanoparticles	Size of nanoparticles (in nm)	Precursors	References
Cynodon dactylon	Bio silica	7 -80	-	Babu et al. (2018)
Croton sparsiflorus	Ag	22–52	AgNO ₃	Kathiravan et al. (2015)
Euphorbia condylocarpa	Pd/ Fe ₃ O ₄	Less than 39	PdCl ₂ & FeCl ₃ 6H ₂ O	Nasrollahzadeh et al. (2015)
Cocos nucifera	Pb	47	Pb(COOH) ₂	Elango and Roopan (2015)
Gloriosa superba	CeO ₂	5	CeCl ₃	Arumugam et al. (2015)
Malva sylvestris	CuO	14	CuCl _{2.} 2H ₂ O	Awwad et al. (2015)
Catharanthus roseus	Pd	38	Pd(OAc)	Kalaiselvi et al. (2015)
Cassia alata	CuO	110-280	$CuSO_4$	Jayalakshmi and Yogamoorthi (2014)
Olea europaea	Ag	20-25	AgNO ₃	Khalil <i>et al.</i> (2014)
Camellia sinensis	ZnO	16	$\frac{\text{Zn}(\text{O}_2\text{CCH}_3)_2(\text{H}_2}{\text{O})_2}$	Senthilkumar and Sivakumar (2014)
Phyllanthus amarus	CuO	20	CuSO ₄	Acharyulu et al. (2014)
Oryza sativa	TiO ₂	13	TiO ₂ (OH) ₂	Ramimoghadam et al. (2014)

Gum karaya	CuO	4.8-7.8	CuCl _{2.} 2H ₂ O	Velora et al. (2013)
<i>Chenopodium album</i> leaf extract	Silver and gold	10-30	AgNO _{3,} Auric acid	Dwivedi and Gopal (2010)
Parthenium hysterophorus leaf	Ag	30-80	AgNO ₃	Parashar et al. (2009)
Mentha piperita leaf	Ag	10-25	AgNO ₃	Parashar et al. (2009)
Capsicum annum	Ag	10-40	-	Li et al. (2007)
Pelargoneum graveolens	Ag	16-40	AgNO ₃	Shankar <i>et al</i> . (2003)
Medicago sativa	Gold	2-20	-	Torresday et al. (2002)

Role of different NPs on Plant Physiology and Biochemistry

In present years, plant interactions with nanoparticles have resulted in a variety of morphological and physiological changes, depending on the properties of the NPs. The chemical composition, size, surface area, reactivity, and dose at which they respond positively determine the adequacy of NPs (Khodakovskaya *et al.*, 2009). This review deals with the possible roles of different types of NPs in seed germination, photosynthesis and plant growth etc.

(i) Silicon dioxide nanoparticles

Lower concentration of SiO₂ (10 & 20 ppm) NPs significantly increased the percentage of seed germination in Sorghum bicolor (Bhatia et al., 2014). Under salt stress, silicon significantly helps in recovery at growth stages, including germination in wheat (Ahmad et al., 1992). By increasing gas exchange and chlorophyll fluorescence parameters such as net photosynthetic rate, transpiration rate, stomatal conductance, PSII potential activity, effective photochemical efficiency, actual photochemical efficiency, electron transport rate, and photochemical quench, nano-SiO₂ promotes plant growth and development (Siddiqui et al., 2014 and Xie et al., 2011). Exogenous application of nano- SiO_2 and nano-titanium dioxide (nano-TiO₂) improves soybean seed germination by increasing nitrate reductase (Lu et al., 2002) and improving the seeds' ability to absorb and utilise water and nutrients (Zheng et al., 2005).

(ii) Zinc Oxide nanoparticles

Studies suggested that zinc oxide NPs induces plant growth and development. Zinc deficiency can lead to disorders in factors regulating growth in plants (Luomg and Kim, 2015). It decreases grain yield, plant protein percentage and the nutritional value of products (Taheri et al.2015). Laware and Raskar (2014) found that at the concentration of 20 and 30µg ml⁻¹ better growth and flowering occurred with higher values of seeded fruit per umbel, seed weight perumbel and 1000 seed weight, it overall reduce the photoperiod in onion. Hafizi and Nasr (2018) found varied concentrations as beneficial nanoparticles on the level of enzymes in safflower. Munir et al. (2018) reported that ZnONPs induced a significant improvement in wheat growth characteristics, photosynthesis and biomass by seed priming method, Zn were found to be in higher concentration in the roots, shoot and grains of wheat than the control reduce the Zn deficiency in plants. Three different physical forms of ZnO particles (ZnO nanocolloid, ZnO nanoparticles, and micrometric ZnO particles)were analyzed in irrigation water supplied to mineral poor soil. Taheri et al. (2015) found that all the three helps in improving shoot dry matter and leaf area index while the best result given by ZnO nanoparticle treatment so they concluded that zinc nanoparticles can

improve corn growth and yield in mineral-depleted soils. Siddiqui et al. (2018) reported that a foliar spray of ZnO NPs to plants lacking Rhizobium resulted in significant improvements in lentil growth, pod number, chlorophyll, carotenoid content, and NR activity in both inoculated and uninoculated plants. Thunugunta et al. (2018) observed positive impact of ZnO nanoparticles to the seedling growth of eggplant under greenhouse conditions while negative impact has been seen under tissue condition. ZnO and ZnSO₄ NPs were examined at 1000 and 2000 mg L^{-1} concentration ZnO NPs at a concentration of 1000 mg L^{-1} positively affected plant height, stem diameter, and chlorophyll content, increased fruit yield and biomass accumulation compared to ZnSO₄ treatments and could be used habanero pepper production to improve yield, quality, and nutraceutical properties of fruits (Lopez et al. 2019)

(iii) Silver Nanoparticles

Salama (2012) studied the effect biologically synthesized AgNPs of different concentrations 20, 40, 60, 80 and 100 ppm on (Phaseolus vulgaris L.) and corn (Zea mays L.) and discovered that biosynthesized AgNPs had a significant impact on the growth of plantlets. Sadak (2019) analyzed the different concentration of AgNPs among the 40mg/l concentration showed the best results in improving the growth parameters and as well in biochemical parameters such as shoot length, number of leaves/plant, shoot dry weight, photosynthetic pigment (chlorophyll a, chlorophyll b, and carotenoids), indole acetic acid (IAA) contents thus enhance the yield quantity (number of pods/plant, number of seeds/pod, weight of seeds/plant, and seed index) quality (carbohydrate%, protein%, phenolics, flavonoids, and tannins contents) of the yielded seeds as well as increasing antioxidant activity of the yielded seeds f fenugreek plant. Recently, Krishnaraj et al. (2012) investigated the effect of biologically synthesised Ag NPs on the growth metabolism of hydroponically grown Bacopa monnieri and concluded that biosynthesized AgNPs have a significant effect on seed germination, induced protein and carbohydrate synthesis, decrease total phenol contents, and catalase and peroxidase activities.

(iv) Titanium dioxide Nanoparticles

Number of studies has been done to understand the impact of titanium dioxide nanoparticles (TiO₂NPs) on bacteria, algae, plankton, fish, mice, and rats but less work has been done on plant (Siddiqui etal. 2015). TiO₂NPs have the capability to affect the food chain, processing and economics of barley (Mattiello and Marchiol, 2017). Dehkourdi andMosavi (2013) reported that parley seeds treated by nano-anatase showed a rise in the percentage of germination, the germination rate index, the root and shoot length, the fresh weight, the vigor index, and the chlorophyll content of the seedlings. TiO₂NPs enhanced seed

germination, radicle and plumule growth of canola seedlings (Mahmoodzadeh et al., 2013). Jiang et al. (2013) studied the combined effects and physiological mechanism of high-CO₂ and TiO₂ NPs on wheat and gives the better understanding of the joint effects to terrestrial plants. Faraji and Sepehri (2018) studied the joint effects of TiO₂ NPs and sodium nitroprusside (SNP) donor of NO (nitric oxide) on seed germination and seedling growth of wheat under cadmium (Cd) stress and concluded that it would be an approach in preventing the negative effects of Cd stress. Both nano titanium dioxide (n-TiO₂) and sodium nitroprusside (SNP)increases tolerance under salinity stressin barley by increasing the some antioxidant enzyme activities (Karami and Sepehri, 2018). Under water deficient stress, foliar application of titanium dioxide nanoparticles increases plant growth characteristics of thyme in Thymus vulgaris (Nasab et al., 2018). TiO₂NPs enhances the nitrate reductase, carbonic anhydrase activities, Chlorophyll fluorescence, net photosynthetic rate, essential oil productivity and yield of Mentha piperita L. (Ahmad et al. 2018).

(v) Gold Nanoparticles

Betwixt metal-based nanoparticles, impact of AuNPs on germination, water balance, nutrition, genotoxicology or seed production is still unexplored (Hendel *et al.*, 2017).Gold nanoparticles are biocompatible and have the potential to be used as nanocarriers in agriculture (Ndeh *et al.*, 2017). Smaller sized gold nanoparticles were more effective in inducing the percentage of seed germination and shoot length of tomato seedlings, and have higher content of chlorophyll in contrast to control plant (Thakur *et al.* 2018).Barrena *et al.* (2009) in lettuce and cucumber, Savithramma *et al.* (2012) in *Boswellia ovalifoliolata*, Arora *et al.* (2012) in *Brassica juncea* and Gopinath *et al.* (2014) in *Gloriosa superba*

observed that AuNPs enhance seed germination. AuNPs enhance the number of leaves, leaf area, plant height, chlorophyll content, and sugar content that lead to the better crop yield.

(vi) Carbon Nanotubes

Betwixt NPs, carbon nanotube (CNTs) possess a significant place due to their exceptional mechanical, electrical, optical and thermal properties (Hurt et al., 2006; Bennett et al., 2013; Srivastava et al., 2015). The majority of current CNTs research has focused on people and animals (Ke et al., 2011 and Tiwari et al., 2014). There has been a scarcity of information on carbon nanotubes and their interactions with plant cells and metabolism (Siddiqui et al., 2015). Two different types of CNTs, single-walled (SWCNT) and multi-walled (MWCNT) (Vithanage et al., 2017). Delivery of chemicals to cells, penetration in the cell wall and cell membrane by carbon nanotube (Siddiqui et al., 2015). CNTs added to soil mix through watering and two times more yield of flower and fruit in comparison to control plant in tomato with regular soil thus act as a plant growth regulator (Khodakovskaya et al., 2013). CNTs greatly influence the cell walls of tomato seed coats and stimulate the seedling growth and germination (Khodakovskaya et al., 2009). Oloumi et al. (2018) observed that the response of MWCNTs on heavy metal and growth parameters accumulation in plant seedlings is mainly depends on heavy metal type, MWCNTs concentration and plant species. Water soluble carbon nanotubes at 6.0 mg mL⁻¹increased growth rate of every part of Cicer arietinum, indicating better water absorption and retention related to enhanced growth (Tripathi et al., 2011). The effectiveness of NPs is determined by their concentration, which varies from plant to plant (Table 2).

Nanoparticles	Effective concentration(s)	Plant	Effect on plant part	References
	400mg/kg	Cucumis sativus fruit	Micronutrients (Cu, Mn, and Zn)	Zhao <i>et al.</i> (2014)
	1.5 ppm (foliar spray)	Cicer arietinum	Improved shoot dry weight	Burman <i>et al.</i> (2013)
ZnO NPs	20 ppm (suspension,foliar spray)	Vigna radiata	Enhanced biomass	Dhoke <i>et al</i> . (2013)
	500, 1,000, 2,000 and 4,000 ppm	Vigna radiate	Increased dry weight	Patra <i>et al.</i> (2013)
	1,000 ppm	Arachis hypogaea	Enhanced stem, root growth and yield	Prasad <i>et al.</i> (2012)
AgNPs	60 ppm	Phaseolus vulgaris, Zea mays	Increased root length	Salama (2012)
	10-30 µg/mL	Boswellia ovalifoliolata	Improved germination and seedling growth	Savithramma et al. (2012)
	60 ppm	Phaseolus vulgaris, Zea mays	Increased Shoot length	Salama (2012)
	60 ppm	Phaseolus vulgaris, Zea mays	Increased Dry weight of root and shoot	Salama (2012)
	100 μM	Vigna radiata	Antagonize inhibition by 2,4 dichlorophenoxyacetic acid (2,4-D) at 500 μM of plant growth	Karuppanapandian <i>et al.</i> (2011)
TiO ₂ NPs	60 ppm	Foenicutum vulgare	Increased germination	Feizi et al. (2013)
	1,000 mg/L	Triticum aestivum	Improved chlorophyll content	Mahmoodzadeh <i>et al.</i> (2013)

Table 2 : Effective role and concentration(s) of Nanoparticles on plant growth and development

	0.05–0.2 g/L	Lycopersicon esculentum	Net photosynthetic rate, conductance to H ₂ O, and transpiration rate, Regulation of photosystem II (PSII)	Qi et al. (2013)
	lower than 200 mg/L	Lemna minor	Enhanced plant growth	Song <i>et al</i> . (2012)
	400 mg/L	Arabidopsis thaliana	Improved root length	Lee et al. (2010)
	0.25 %	Spinacia oleracea	Hill reaction, non-cyclic photophosphorylation, protect chloroplasts from aging	Hong <i>et al</i> . (2005a, b)
SiO ₂ NPs	20ppm	Sorghum bicolor	Seedling biology	Bhatia <i>et al.</i> (2014)
510 ₂ 1 NF 8	15 kg/ha	Zea mays	Improved growth parameters	Yuvakkumar et al. (2011)
Iron oxide NPs	50 ppm (foliar spray)	Vigna radiata	Increased Biomass	Dhoke <i>et al.</i> (2013)
from oxide NPS	0.5–0.75 g/L	Glycine max	Enhanced yield and quality	Sheykhbaglou et al. (2010)
MWCNTs	50-200µg/ml	Lycopersicum esculentum	Improvement in germination and seedling growth	Khodakovskaya <i>et al.</i> (2013)

Conclusion and future perspective

Due to unique properties, nanoparticles have become an important topic of research in recent years such as agriculture, health care, environment, etc. This review displays the effect of various nanoparticles on different plants of many concentrations, sizes, and shapes. It is evident from compiled knowledge that the effect of NPs is distinct from plant to plant. Future research should focus on molecular/genetic response in the presence of NPs, mode of action of NPs and the interaction with biomolecules. In Green synthesis, numerous kinds of natural resources have been used viz. yeast, bacteria, fungi and plant extract, plant extract among all has proven the best as stabilizing and reducing agent.

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References

- Acharyulu, N.P.S.; Dubey, R.S.; Swaminadham, V.; Kalyani, R.L.; Kolluand, P. and Pammi, S.V.N. (2014). Green synthesis of Cuo nanoparticles using *Phyllanthus amarus* leaf extract and their antibacterial activity against multidrug resistance bacteria. *Int. J.Eng. Res. Technol.* 3: 639-641.
- Agarwal, H.; Kumar, S.V. and RajeshKumar, S. (2017). A review on green synthesis of zinc oxide nanoparticles– An eco-friendly approach. *Res.-Effi. Technol.* 3: 406-413.
- Ahmad, B.; Shabbir, A.; Jaleel, H.; Khan, M.M.A. and Sadiq, Y. (2018). Efficacy of titanium dioxide nanoparticles in modulating photosynthesis, peltate glandular trichomes and essential oil production and quality in *Mentha piperita* L. *Curr.Plant Bio.*, 13: 6–15.
- Ahmad, R.; Zaheer, S.H. and Ismail, S. (1992). Role of silicon in salt tolerance of wheat (*Triticum aestivum* L.), *Plant Sci.*, 85: 43-50.
- Arora, S.; Rajwade, J.M. and Paknikar, K.M. (2012). Nanotoxicology and *in vitro* studies: The need of the hour. *Toxicol. and App. Pharmacol.*, 258: 151–165.
- Arora, S.; Sharma, P.; Kumar, S.; Nayan, R.; Khanna, P.K. and Zaidi, M.G.H. (2012). Gold-nanoparticle induced

enhancement in growth and seed yield of *Brassica* juncea. Plant Growth Regul. 66: 303–310.

- Arumugama, A.; Karthikeyan, C.; Hameed, A.S.H.; Gopinath, K.; Gowri, S. and Karthika, V. (2015). Synthesis of cerium oxide nanoparticles using *Gloriosa superba* L. leaf extract and their structural, optical and antibacterial properties. *Mater.Sci. Eng.* 49: 408–415.
- Awwad, A.M.; Albiss, B.A. and Salem, N.M. (2015). Antibacterial activity of synthesized copper oxide nanoparticles using *Malva sylvestrism* leaf extract. *SMU Med. J.* 2: 91-101.
- Nair, B. and Pradeep, T. (2002). Coalescence of nanoclusters and formation of submicron crystallites assisted by *Lactobacillus* Strains cryst. *Growth Des.* 2: 293–298.
- Babu, R.H.; Yugandhar, P. and Savithramma, N. (2018). Synthesis, characterization and antimicrobial studies of bio silica nanoparticles prepared from *Cynodon dactylon* L.: A green approach. *Bull.Mater. Sci.*, 41: 1-8.
- Barrena, R.; Casals, E.; Colon, J.; Font, X.; Sanchez, A. and Puntes, V. (2009). Evaluation of the ecotoxicity of model nanoparticles. *Chemosphere*, 75: 850–857.
- Bennett, S.W.; Adeleye, A.; Jiand, Z. and Keller, A.A. (2013). Stability, metal leaching, photoactivity and toxicity in freshwater systems of commercial single wall carbon nanotubes. *Water Res.*, 47: 1-12.
- Bhatia, S.S.; Bahri, S. and Moitra, S. (2014). SiO₂ nanoparticles: effect on seedling biology. *Int. J. App. Eng. Res.* 9: 935-939.
- Burman, U.; Saini, M. and Kumar, P. (2013). Effect of zinc oxide nanoparticles on growth and antioxidant system of chickpea seedlings. *Toxicol. Environ. Chem.*, 95: 605–612.
- Chung, I.L.; Park, I.; Hyun, K.S.; Thiruvengadam, M. and Rajakumar, G. (2016). Plant-mediated synthesis of silver nanoparticles: their characteristic properties and therapeutic applications. *Nanoscale Res. Lett.* 11:1-14.
- Dehkourdi, E.H. and Mosavi, M. (2013). Effect of anatase nanoparticles (TiO₂) on Parsley seed germination (*Petroselinum crispum*) in Vitro. *Biol. Trace Elem. Res.*, 155: 283–286.
- Dhoke, S.K.; Mahajan, P.; Kamble, R. and Khanna, A. (2013). Effect of nanoparticles suspension on the growth of mung (*Vigna radiata*) seedlings by foliar spray method. *Nanotechnol. Dev.* 3: e1.

- Dwivedi, A.D. and Gopal, K. (2010). Biosynthesis of silver and gold nanoparticles using *Chrenopodium album* leaf extract. Colloids surf.: *Physiochem. & Eng. Asp.*, 369: 27-33.
- Elango, G. and Roopan, S.M. (2015). Green synthesis, spectroscopic investigation and photocatalytic activity of lead nanoparticles. Spectrochim. *Acta A: Mol. Biomol. Spectrosc*, 139: 367–373.
- Faraji, J. and Sepehri, A. (2018). Titanium dioxide nanoparticles and sodium nitroprusside alleviate the adverse effects of cadmium stress on germination and seedling growth of Wheat (*Triticum aestivum* L.). Univ. Sci., 23: 61-87.
- Feizi, H.; Kamali, M.; Jafari, L. and Moghaddam, P.R. (2013). Phytotoxicity and stimulatory impacts of nanosized and bulk titanium dioxide on fennel (*Foeniculum vulgare* Mill). *Chemosphere*, 91: 506–511.
- Galbraith, D.W. (2007). Nanobiotechnology: silica breaks through in plants. *Nat. Nanotechnol.* 2: 272–273.
- Giraldo, J.P.; Landry, M.P.; Faltermeier, S.M.; McNicholas, T.P.; Iverson, N.M.; Boghossian, A.A.; Reuel, N.F.; Hilmer, A.J.; Sen, F.; Brew, J.A. and Strano, M.S. (2014). Plant nanobionics approach to augment photosynthesis and biochemical sensing. *Nat. Mater.* 13: 400-8.
- Gopinath, K.; Gowri, S.; Karthika, V. and Arumugam, A. (2014). Green synthesis of gold nanoparticles from fruit extract of *Terminalia arjuna*, for the enhanced seed germination activity ofGloriosa superb. J. Nanostruct Chem. 4: 1–11.
- Gutierrez, F.J.; Mussons, M.L.; Gaton, P. and Rojo, R. (2011). Nanotechnology and food industry. Scientific, health and social aspects of the food industry, pp. 95-128
- Hafizi, Z. and Nasr, N. (2018). The effect of zinc oxide nanoparticles on safflower plant growth and physiology. *Eng. Technol App. Sci. Res.*, 8: 2508-2513.
- Hendel, A.M.; Zubko, M.; Karcz, J.; Strozand, D. and Kurczynska, E. (2017). Fate of neutral-charged gold nanoparticles in the roots of the *Hordeum vulgare* L. *cultivar Karat.Sci. Rep.*, 7: 3014.
- Hong, F.; Zhou, J.; Liu, C.; Yang, F.; Wu, C.; Zheng, L. and Yang, P. (2005a). Effect of nano-TiO₂ on photochemical reaction of chloroplasts of spinach, *Biol. Trace Elem. Res.*, 105: 269–279.
- Hong, F.S.; Yang, F.; Ma, Z.N.; Zhou, J.; Liu, C.; Wu, C. and Yang, P. (2005b). Influences of nano-TiO₂ on the chloroplast ageing of spinach under light. *Biol. Trace Elem. Res.*, 104: 249–260.
- Hurt, R.H.; Monthioux, M. and Kane, A. (2006). Toxicology of carbon nanomaterials: status, trends, and perspectives on the special issue. *Carbon* 44: 1028–1033.
- Jayalakshmi, Y.A. and Yogamoorthi, A. (2014). Green synthesis of copper oxide nanoparticles using aqueous extract of *Cassia alata* and particles characterization, *Int. J. Nanomat. Biostuct.* 4: 66-71.
- Jiang, F.; Shen, Y.; Ma, C.; Zhang, X.; Cao, W. and Rui, Y. (2017). Effects of TiO₂ nanoparticles on Wheat (*Triticum aestivum L.*) seedlings cultivated under superelevated and normal CO₂ conditions. Plos One 12: e0178088.
- Kalaiselvi, A.; Roopan, S.M.; Madhumitha, G.; Ramalingam,C. and Elango, G. (2015). Synthesis and characterization of palladium nanoparticles using

Catharanthus roseus leaf extract and its application in the photo-catalytic degradation. Spectrochim. *Acta A: Mol. Biomol. Spectrosc.*135: 116–119.

- Kalpana, V.N. and Rajeswari, V.D. (2018). A review on green synthesis, biomedical applications, and toxicity studies of ZnO NPs. *Bioinorg. Chemist. App.*, 2018: 1-12.
- Karami, A. and Sepehri, A. (2018). Nano titanium dioxide and nitric oxide alleviate salt induced changes in seedling growth, physiological and photosynthesis attributes of Barley, *Zemdirbyste-Agri*. 105: 123-132.
- Karuppanapandian, T.; Wang, H.W.; Prabakaran, N.; Jeyalakshmi, K.; Kwon, M.; Manoharan, K. and Kim, W. (2011). 2,4-dichlorophenoxyacetic acid-induced leaf senescence in Mung bean (*Vignaradiata* L. Wilczek) and senescence inhibition by co-treatment with silver nanoparticles. *Plant Physiol. Biochem.* 49: 168–177.
- Kathiravan, V.; Ravi, S.; Ashokkumar, S.; Velmurugan, S.; Elumalai, K. and Khatiwada, C.P. (2015). Green synthesis of silver nanoparticles using *Croton* sparsiflorus morong leaf extract and their antibacterial and antifungal activities. Spectrochim. Acta A: Mol. Biomol. Spectrosc., 139: 200–205.
- Kaushik, N.; Thakkar, M.S.; Snehit, S.; Mhatre, M.S.; Rasesh, Y.; Parikh, M.S. (2010). Biological synthesis of metallic nanoparticles. *Nanomed Nanotechnol Biol. Med.*, 6: 257–262.
- Ke, P.C.; Lin, S.; Reppert, J.; Rao, A.M. and Luo, H. (2011). Uptake of carbon-based nanoparticles by mammalian cells and plants. In: Handbook of nanophysics: nanomedicine and nanorobotics, K.D. Sattler (eds), CRC Press, New York, pp. 1–30.
- Khalil, M.M.H.; Ismail, E.H.; Baghdady, K.Z.E. and Mohamed, D. (2014). Green synthesis of silver nanoparticles using olive leaf extract and its antibacterial activity. *Arab.J. Chem.*, 7: 1131–1139.
- Kharissova, O.V.; Dias, H.V.R.; Kharisov, B.I.; Perez, B.O.; Victor, M. and Perez, J. (2013). The greener synthesis of nanoparticles. *Trends Biotechnol.*, 31: 240–248.
- Khodakovskaya, M.; Dervishi, E.; Mahmood, M.; Xu, Y.; Li, Z.; Watanabe, F. and Biris, A.S. (2009). Carbon nanotubes are able to penetrate plant seed coat and dramatically affect seed germination and plant growth, ACS Nano 3: 3221–3227.
- Khodakovskaya, M.V.; Kim, B.; Kim, J.N.; Alimohammadi, M.; Dervishi, E.; Mustafa, T. and Cernigla, C.E. (2013). Carbon nanotubes as plant growth regulators: effects on tomato growth, reproductive system, and soil microbial community. Small 9: 115-123.
- Krishnaraj, C.; Jagan, E.G.; Ramachandran, R.; Abirami, S.M.; Mohan, N. and Kalaichelvan, P.T. (2012). Effect of biologically synthesized silver nanoparticles on *Bacopa monnieri* (Linn.)Wettst.plant growth metabolism. *Process Biochem*, 47: 51–658.
- Landry, M.P.; Faltermeier, S.M.; McNicholas, T.P.; Iverson, N.M.; Boghossian, A.A.; Reuel, N.F.; Hilmer, A.J.; Sen, F.; Brew, J.A. and Strano, M.S. (2014). Plant nanobionics approach to augment photosynthesis and biochemical sensing. *Nat. Mater*, 13: 400–408.
- Laware, S.L. and Raskar, S. (2014). Influence of zinc oxide nanoparticles on growth, flowering and seed productivity in Onion. *Int. J. Curr. Microbiol. App. Sci.*, 3: 874-881.

- Lee, C.W.; Mahendra, S.; Zodrow, K.; Li, D.; Tsai, Y.C.; Braam, J. and Alvarez, P.J.J. (2010). Developmental phytotoxicity of metal oxide nanoparticles to *Arabidopsis thaliana. Environ. Toxicol. Chem.* 29: 669– 675.
- Leon, E.R.; Palomares, R.I.; Navarro, R.E.; Urbina, R.H.; Tanori, J.; Palomaresand, C.I. and Maldonado, A. (2013). Synthesis of silver nanoparticles using reducing agents obtained from natural sources (*Rumex hymenosepalus* extracts). *Nanoscale Res. Lett.* 8: 318.
- Lopez, J.I.G.; Medina, G.N.; Saenz, E.O.; Saldivar, R.H.L.; Castro, E.D.B.; Alvarado, R.V.; Salinasand, P.A.R. and Garcia, F.Z. (2019). Foliar application of zinc oxide nanoparticles and zinc sulfate boosts the content of bioactive compounds in Habanero peppers. *Plants* 8: 254.
- Lu, C.M.; Zhang, C.Y.; Wen, J.Q.; Wu, G.R. and Tao, M.X. (2002). Research on the effect of nanometer materials on germination and growth enhancement of *Glycine max* and its mechanism. *Soybean Sci.* 21: 68–172.
- Luomg, B. and Kim, J. (2015). Synthesis & characterization of nano sized Zn powder by electrical explosion. World scientific 28 octobr 3903-3909.
- Mahmoodzadeh, H.; Nabavi, M. and Kashefi, H. (2013). Effect of nanoscale titanium dioxide particles on the germination and growth of canola (*Brassica napus*). J. *Ornam. Hortic. Plants* 3: 25-32.
- Mattiello, A.; Marchiol, L. (2017). Application of nanotechnology in agriculture: assessment of TiO₂ nanoparticle effects on Barley. In: Application of Titanium Dioxide, M., Janus (eds), InTech: London, UK, pp. 23-39.
- Meyers, M.A.; Mishra, A. and Benson, D.J. (2006). Mechanical properties of nanocrystalline materials. *Prog. Mater Sci.* 51: 427–556.
- Mishra, V. and Sharma, R. (2015). Green synthesis of zinc oxide nanoparticles using fresh peels extract of *Punica* granatum and its antimicrobial activities. *Int. J. Pharma. Res. and Health Sci.*, 3: 694–699.
- Mukherjee, P.; Ahmad, A.; Mandal, D.; Senapati, S.; Sainkara, S.R. and Khan, M.I. (2001). Fungus mediated synthesis of silver nanoparticles and their immobilization in the mycelial matrix: A novel biological approach to nanoparticle synthesis. *Nano Lett.* 1: 515–519.
- Munir, T.; Rizwan, M.; Kashif, M.; Shahzad, A.; Alib, S.; Amin, N.; Zahid, R.; Alam, M.F.E. and Imran, M. (2018). Effect of zinc oxide nanoparticles on the growth and Zn uptake in Wheat (*Triticum aestivum* L.) by seed priming method. *Dig. J. Nanomater. Biostruct.* 13:315-323.
- Nair, R.; Varghese, S.H.; Nair, B.G.; Maekawa, T.; Yoshida, Y.; Kumar, D.S. (2010). Nanoparticulate material delivery to plants. *Plant Sci.* 179: 154–163.
- Nasab, B.F.; Sirousmehrand, A.R. and Azad, H. (2018). Effect of titanium dioxide nanoparticles on essential oil quantity and quality in *Thymus vulgaris* under water deficit. J. Med. Plants by-products 2: 125-133.
- Nasrollahzadeha, M.; Sajadib, S.M.; Vartoonia, A.R. and Khalajc, M. (2015). Green synthesis of Pd/Fe₃O₄ nanoparticles using *Euphorbia condylocarpa* M. bieb root extract and their catalytic applications as magnetically recoverable and stable recyclablecatalysts

for the phosphine-free Sonogashira and Suzuki coupling reactions, *J. Mol. Catal. A Chem.* 396: 31–39.

- Ndeh, N.T.; Maensiriand, S. and Maensiri, D. (2017). The effect of green synthesized gold nanoparticles on rice germination and roots. *Adv. Nat. Sci.: Nanosci. Nanotechnol.* 8: 1-10.
- Nel, A.; Xia, T.; Madler, L. and Li, N. (2006). Toxic potential of materials at the nanolevel. *Sci.* 311: 622–7.
- Oloumi, H.; Mousavi, E.A. and Nejad, R.M. (2018). Multiwall carbon nanotubes effects on plant seedlings growth and cadmium/lead uptake in vitro. *Russ. J. Plant Physiol.* 65: 260–268.
- Parashar, U.K.; Saxena, P.S. and Srivastava, A. (2009). Bioinspired synthesis of silver nanoparticles. *Dig. J. Nanomater. Biostruct.* 4: 159 – 166.
- Parashar, V.; Parashar, R.; Sharma, B. and Pandey, A.C. (2009).*Parthenium* leaf extract mediated synthesis of silver nanoparticles: A novel approach towards weed utilization. *Mat. Sci.* 4: 45-50.
- Patra, P.; Choudhury, S.R.; Mandal, S.; Basu, A.; Goswami, A.; Gogoi, R.; Srivastava, C.; Kumar, R. and Gopal, M. (2013). Effect sulfur and ZnO nanoparticles on stress physiology and plant (*Vigna radiata*) nutrition. In: Advanced Nanomaterials and Nanotechnology, P.K. Giri *et al.* (eds), *Springer Berlin Heidelberg*, pp. 301-309.
- Prasad, T.N.V.K.V.; Sudhakar, P.; Sreenivasulu, Y.; Latha, P.; Munaswamy, V.; Raja Reddy, K.; Sreeprasad, T.S.; Sajanlaland, P.R. and Pradeep, T. (2012). Effect of nanoscale zinc oxide particles on the germination, growth and yield of peanut. J. Plant Nutr. 35: 905–927.
- Qi, M.; Liu, Y. and Li, T. (2013). Nano-TiO₂ improve the photosynthesis of tomato leaves under mild heat stress. *Biol. Trace. Elem. Res.* 156: 323–328.
- Ramimoghadam, D.; Bagheri, S.; Bee, S. and Hamid, A. (2014). Biotemplated synthesis of anatase titanium dioxide nanoparticles via lignocellulosic waste material. *BioMed Res. Int.* 2014: 1-7.
- Sadak, M.S. (2019). Impact of silver nanoparticles on plant growth, some biochemical aspects and yield of fenugreek plant (*Trigonella foenum-graecum*). *Bull. Natl. Res. Cent.* 43: 1-6.
- Salama, H.M.H. (2012). Effects of silver nanoparticles in some crop plants, common bean (*Phaseolus vulgaris* L.) andcorn (*Zea mays* L.). *Int. Res. J. Biotech.* 3: 190– 197.
- Savithramma, N.; Ankanna, S. and Bhumi, G. (2012). Effect of nanoparticles on seed germination and seedling growth of *Boswellia ovalifoliolata* an endemic and endangered medicinal tree taxon. *Nano Vision*, 2: 61– 68.
- Schultz, S.; Smith, D.R.; Mock, J.J. and Schultz, D.A. (2000). Single-target molecule detection with nonbleaching multicolor optical immunolabels, *Proc. Natl. Acad. Sci. USA* 97: 996–1001.
- Senthilkumar, S.R. and Sivakumar, T. (2014). Green tea (*Camellia sinensis*) mediated synthesis of zinc oxide (ZnO) nanoparticles and studies on their antimicrobial activities, *Int. J. Pharm. Sci.* 6: 461-465.
- Sepeur, S. (2008). Nanotechnology: technical basics and applications, Vincentz Network GmbH & Co KG.
- Shankar, S.S.; Ahmad, A. and Sastry, M. (2003). Geranium leaf assisted biosynthesis of silver nanoparticles. *Biotechnol. Prog.* 19: 1627-1631.

- Sheykhbaglou, R.; Sedghi, M.; Shishevan, M.T. and Sharifi, R.S. (2010). Effects of nano-iron oxide particles on agronomic traits of soybean. *Not. Sci. Biol.*, 2: 112–113.
- Shikuo, L.; Yuhua, S.; Anjian, X.; Xuerong, Y.; Lingguang, Q.; Li, Z. and Zhang, Q. (2007). Green synthesis of silver nanoparticles using *Capsicum annuum* L extract. *Green Chem.* 9: 852-858.
- Siddiqui, M.H.; Al-Whaibi, M.H.; Faisal, M. and Al-Sahli, A.A. (2014). Nano-silicon dioxide mitigates theadverse effects of salt stress on *Cucurbita pepo L. Environ. Toxicol. Chem.*, 33: 2429–2437.
- Siddiqui, M.H.; Al-Whaibi, M.H.; Firoz, M. and Al-Khaishany, M.Y. (2015). Role of nanoparticles in plants. In: Nanotechnology and Plant Sciences, M. Siddiqui, Al-Whaibi M., Mohammad F. (eds). Springer, Cham, pp. 19-35.
- Siddiqui, Z.A.; Khan, A.; Khan, M.R. and Abd-Allah, E.F. (2018). Effects of zinc oxide nanoparticles (ZnO NPs) and some plant pathogens on the growth and nodulation of lentil (*Lens culinaris* Medik). *Acta Phytopathol. Entomol. Hung.* 53: 195–212.
- Singh, A.; Singha, N.B.; Hussaina, I.; Singh, H. and Singh, S.C. (2015). Plant-nanoparticle interaction: An approach to improve agricultural practices and plant productivity. *Int. J. Pharma. Sci. Inven.*, 4: 25-40.
- Song, G.; Gao, Y.; Wu, H.; Hou, W.; Zhang, C. and Ma, H. (2012). Physiological effect of anatase TiO₂ nanoparticles on *Lemna minor. Environ. Toxicol. Chem.* 31: 2147–2152.
- Srivastava, V.; Gusain, D. and Sharma, Y.C. (2015). Critical review on the toxicity of some widely used engineered nanoparticles. *Ind. Eng. Chem. Res.*, 54: 6209–6233.
- Taheri, M.; Qarache, H.A.; Qarache, A.A. and Yoosefi, M. (2015). The effects of zinc-oxide nanoparticles on growth parameters of Corn (SC704), STEM Fellowship J. 1: 17-20.
- Thakkar, K.N.; Mhatre, S.S. and Parikh, R.Y. (2010). Biological synthesis of metallic nanoparticles. *Nanomed Nanotechnol. Biol. and Med.*, 6: 257–262.
- Thakur, R.K.; Dhirtaand, B. and Shirkot, P. (2018). Studies on effect of gold nanoparticles on *Meloidogyne incognita* and tomato plants growth and development. *Bio. Rxiv.*, 1: 428144.
- Thunugunta, T.; Reddy, A.C.; Seetharamaiah, S.K.; Hunashikatti, L.R.; Chandrappa, S.G. and Kalathil, N.C. (2018). Reddy Impact of Zinc oxide nanoparticles on eggplant (*S. melongena*): studies on growth and the

accumulation of nanoparticles. *IET Nanobiotechnol.* 12: 706-713.

- Tiwari, D.K.; Dasgupta-Schubert, N.; Villasenor-Cendejas, L.M.; Villegas, J.; Carreto-Montoya, L. and Borjas-Garcia, S.E. (2014).Interfacing carbon nanotubes (CNT) with plants: Enhancement of growth, water and ionic nutrient uptake in maize (*Zea Mays*) and implications for nanoagriculture. *Appl. Nanosci.* 4: 577–591.
- Torney, F.; Trewyn, B.G.; Lin, V.S. and Wang, K. (2007). Mesoporous silica nanoparticles deliver DNA and chemicals into plants. *Nat. Nanotechnol.* 2: 295–300.
- Torresday, J.L.G.; Parsons, J.G.; Gomez, E.; Videa, J.P.; Troiani, H.E.; Santiago, P. and Yacaman, M.J. (2002). Formation and growth of Au nanoparticles inside live AlfaAlfa plants. *Nano letters.*, 2: 397-401.
- Tripathi, S.; Sonkar, S.K. and Sarka, S. (2011). Growth stimulation of gram (*Cicer arietinum*) plant by water soluble carbon nanotubes. *Nanoscale*, 3: 1176-1181.
- Vellora, V.; Padil, T. and Cerník, M. (2013). Green synthesis of copper oxide nanoparticles using *Gum karaya* as a biotemplate and their antibacterial application. *Int. J. Nanomed.*, 8: 889–898.
- Vithanage, M.; Seneviratne, M.; Ahmad, M. and Sarkarand, Y.S. (2017). Contrasting effects of engineered carbon nanotubes on plants: a review. *Environ. Geochem. Health*, 39: 1421-1439.
- Willner, I.; Baron, R. and Willner, B. (2006). Growing metal nanoparticles by enzyme, *Adv. Mater.*, 18: 1109–1120.
- Xie, Y.; Li, B.; Zhang, Q.; Zhang, C.; Lu, K. and Tao, G. (2011). Effects of nano-TiO₂ on photosynthetic characteristics of *Indocalamus barbatus*. J. Northeast Univ. 39: 22–25.
- Yuvakkumar, R.; Elango, V.; Rajendran, V.; Kannan, N.S. and Prabu, P. (2011). Influence of nanosilica powder on the growth of Maize crop (*Zea mays L.*). *Int. J. Green Nanotechnol.* 3: 80–190.
- Zhao, L.; Peralta-Videa, J.R.; Rico, C.M.; Hernandez-Viezcas, J.A.; Sun, Y.; Niu, G.; Duarte-Gardea, M. and Gardea-Torresdey, J.L. (2014). CeO₂ and ZnO nanoparticles change the nutritional qualities of cucumber (*Cucumis sativus*). J. Agricul. Food Chem. 62: 2752–2759.
- Zheng, L.; Hong, F.; Lu, S. and Liu, C. (2005). Effect of nano-TiO₂ on strength of naturally aged seeds and growth of spinach. *Biol. Trace Elem. Res.*, 104: 83–91.